

# Montelukast, a leukotriene receptor antagonist, reduces the concentration of leukotrienes in the respiratory tract of children with persistent asthma

Benjamin Volovitz, MD,<sup>a,b</sup> Elvan Tabachnik, MD,<sup>c</sup> Moshe Nussinovitch, MD,<sup>b</sup> Biana Shtaif, MSc,<sup>b</sup> Hanna Blau, MD,<sup>a</sup> Irit Gil-Ad, PhD,<sup>b</sup> Abraham Weizman, MD,<sup>b</sup> and Itzhak Varsano, MD<sup>a,b</sup> *Petah Tikva, Tel Aviv, and Rehovot, Israel*

**Background:** Leukotrienes are bronchoactive mediators secreted by inflammatory cells in the respiratory mucosa on exposure to asthma triggers.

**Objective:** We investigated the effect of montelukast, a leukotriene receptor antagonist, on the release of leukotrienes in the respiratory mucosa of children with persistent asthma.

**Method:** Twenty-three children aged 6 to 11 years with moderately severe asthma were treated in a cross-over design starting, after a 2-week run-in period, with either montelukast (n = 12) or cromolyn (n = 11) for 4 weeks with a 2-week washout period between treatments. Twelve of them were then treated with either montelukast or beclomethasone for 6 months. The use of  $\beta_2$ -agonists was recorded on a diary card. The concentration of leukotriene C<sub>4</sub> (LTC<sub>4</sub>) was measured by HPLC in nasal washes obtained before and at the end of each treatment period. Eosinophilic cationic protein (ECP) was measured in the nasal washes by RIA.

**Results:** The LTC<sub>4</sub> concentration significantly decreased in the children treated for the first 4 weeks with montelukast, from  $5.03 \pm 1.17$  to  $1.42 \pm 0.33$  ng/mL ( $P < .005$ ), and a nonsignificant increase was noted in children treated with cromolyn, from  $3.37 \pm 1.11$  to  $5.88 \pm 2.17$  ng/mL ( $P = .17$ ). ECP concentration also decreased in the children receiving montelukast ( $P = .12$ ). The concentration of LTC<sub>4</sub> remained low after 3 and 6 months of treatment with montelukast ( $0.8 \pm 0.7$  and  $1.0 \pm 0.3$   $\mu$ g/mL) and was lower than with beclomethasone. Children treated with montelukast required significantly fewer  $\beta_2$ -agonists ( $P < .04$ ).

**Conclusion:** Montelukast reduces the concentration of leukotrienes in the respiratory tract of children with persistent asthma parallel to reduction in ECP and clinical improvement. This effect was not observed when the same children were treated with cromolyn. (*J Allergy Clin Immunol* 1999;104:1162-7.)

**Key words:** Leukotrienes, nasal washes, asthma, children, montelukast, leukotriene modifiers

## Abbreviations used

BAL: Bronchoalveolar lavage  
CysLT<sub>1</sub>: Cysteinyl leukotriene 1 (receptor)  
ECP: Eosinophilic cationic protein  
LTC<sub>4</sub>: Leukotriene C<sub>4</sub>  
LTD<sub>4</sub>: Leukotriene D<sub>4</sub>  
LTE<sub>4</sub>: Leukotriene E<sub>4</sub>

Cysteinyl leukotrienes are potent proinflammatory mediators produced from a variety of inflammatory cells, including mast cells, eosinophils, basophils and macrophages. Leukotriene C<sub>4</sub> (LTC<sub>4</sub>) is metabolized enzymatically to leukotriene D<sub>4</sub> (LTD<sub>4</sub>) and subsequently to leukotriene E<sub>4</sub> (LTE<sub>4</sub>), which is excreted in the urine.<sup>1</sup> The leukotrienes have been implicated in the pathophysiologic mechanisms of asthma.<sup>2</sup> LTE<sub>4</sub> has been detected mainly in the urine<sup>3-5</sup> but also in the blood<sup>6,7</sup> and nasal<sup>8</sup> and bronchoalveolar lavage (BAL) fluid<sup>9</sup> of patients with asthma,<sup>9,10</sup> especially after allergen challenge,<sup>4,6,8</sup> after exercise,<sup>3,11</sup> or during an acute asthma attack.<sup>12</sup> LTC<sub>4</sub><sup>8,13-19</sup> and, in lesser amounts, LTD<sub>4</sub><sup>14</sup> have also been detected in the blood<sup>20</sup> and in nasal,<sup>8,15-18,21</sup> tracheal,<sup>18</sup> and BAL fluid<sup>13,14,19</sup> of symptomatic patients with asthma<sup>13,20</sup> and after natural exposure to allergens,<sup>17</sup> viral infection,<sup>15,16</sup> and antigen challenge.<sup>8,14</sup>

Montelukast is a leukotriene receptor antagonist that has been found to be effective in the treatment of asthma in adults<sup>22</sup> and children.<sup>23</sup> There are no data on the effect of montelukast on the presence of leukotrienes in the human respiratory tract.

The current study compares the concentration of leukotrienes in nasal washes of children with persistent asthma treated in a cross-over design with montelukast and cromolyn. The concentration of eosinophilic cationic protein (ECP) in the nasal washes was also measured to determine the relationship of eosinophils to leukotriene concentration.

## METHODS

### Patient population

The study population consisted of 26 children with asthma aged 6 to 11 years, with FEV<sub>1</sub> between 60% to 85% predicted and improvement of  $\geq 12\%$  after inhaled  $\beta_2$ -agonist. During the 2 weeks period before inclusion in the run-in and 2 weeks during run-in the

From the <sup>a</sup>Schneider Children's Medical Center of Israel and the <sup>b</sup>Felsenstein Research Laboratory, Petah Tikva, Sackler School of Medicine, Tel Aviv University, Tel Aviv, and <sup>c</sup>Kaplan Hospital, Rehovot, Israel.

Supported in part by Merck.

Received for publication Mar 18, 1999; revised July 29, 1999; accepted for publication July 29, 1999.

Reprint requests: Benjamin Volovitz, MD, Asthma Clinic, Department of Pediatrics C, Schneider Children's Medical Center of Israel, 14 Kaplan St, Petah Tikva 49202, Israel.

Copyright © 1999 by Mosby, Inc.

0091-6749/99 \$8.00 + 0 1/1/101985

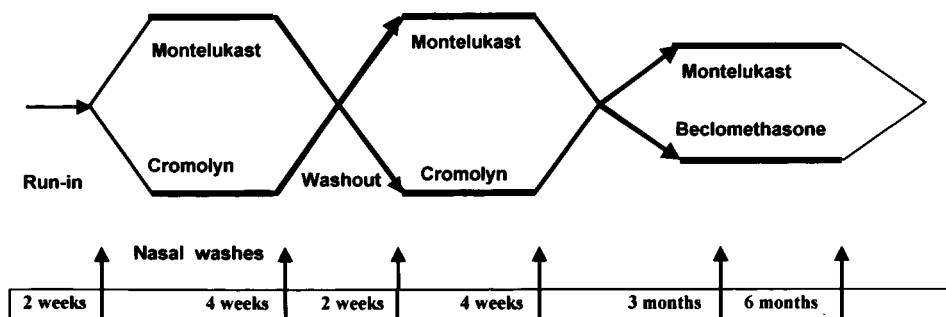


FIG 1. Study design.

TABLE I. Study population: baseline data

Group	Montelukast→ cromolyn	Cromolyn→ montelukast	Statistical significance
No. of children	12	11	
Mean age	9 y 3 mo	8 y 8mo	NS*
Sex (male/female)	6/6 (50%)	8/3 (73%)	NS†
Asthma in family	10/2 (83%)	9/2 (82%)	NS†
Previous inhaled steroid therapy	10/2 (83%)	7/4 (64%)	NS†
FEV <sub>1</sub> (% of predicted) before run-in period	73	78	NS*
FEV <sub>1</sub> (% of predicted) after run-in period	71	75	NS*

NS, Not significant.

\*One-way ANOVA.

†Fisher's exact test.

children were not allowed to use any antiasthma medication except for  $\beta_2$ -agonist on an as-needed basis. During the entire study the children were not permitted to use any nasal drugs. Only children who required  $\beta_2$ -agonists on at least 7 of the 14 days during the run-in period were included in the study. The Helsinki committee of our hospital approved the study. The goals and risks of the study were explained to the parents, and signed informed consent forms were obtained.

### Study design

An open-label, 2-period cross-over design was used. After a 2-week run-in period patients received either montelukast (one 5-mg chewable tablet at bedtime) for 4 weeks followed by cromolyn (2 puffs of 1 mg cromolyn four times daily by metered-dose inhaler) for 4 weeks or cromolyn followed by montelukast, with a 2-week washout period between treatments (Fig 1).  $\beta_2$ -Agonists were used as needed, and their use was recorded on a daily diary card. The children were included in a 6-month extension study wherein they were randomly assigned to receive either montelukast or inhaled beclomethasone (100  $\mu$ g 3 times daily) (Fig 1).

### Sample collection

Nasal fluid washes were collected from each patient on 4 occasions: (1) before onset of the trial (end of run-in period), (2) at the end of the first 4-week treatment period with montelukast or cromolyn, (3) at the end of the 2-week washout period, and (4) at the end of the second 4-week treatment period with the other drug. They were also collected from the children in the extension study after 3 and 6 months of treatment. The samples were collected by suction catheter and a trap. A 4-mL volume of PBS was gently instilled into the nostrils in 8 consecutive 0.5-mL installations, followed each time by gentle suction. The suction catheter was rinsed with 0.5 mL of PBS. Samples were kept on ice during transport to the laboratory and were stored at  $-70^\circ\text{C}$  until assayed.

### Measurement of leukotrienes

The nasal washes containing the secretions in PBS were mixed with 4 volumes of methanol (HPLC grade). After incubation in ice for 2 hours, the samples were centrifuged at 2000 revolutions/min for 10 minutes at  $4^\circ\text{C}$ . The supernatant was applied to a Sep-Pak C<sub>18</sub> cartridge (Millipore, Waters Associates, Milford, Mass) previously activated with 10 mL of 0.5% EDTA, 10 mL of methanol, and 10 mL of water and then eluted with 5 mL of 80% methanol and 20% water (both HPLC grade). This sample was subjected to vacuum extraction to dryness with use of a new small plastic flask (Azlon) for each sample, and the residue was redissolved in 0.75 mL of 30% methanol. A 0.5-mL sample was injected into a C<sub>18</sub> reversed-phase column (Beckman, San Ramon, Calif) with 2 ng of purified PGB<sub>2</sub> (Sigma Chemical, St Louis, Mo) added as an internal standard. A second sample of 0.2 mL was tested whenever the result of the first sample was technically uncertain. LTC<sub>4</sub>, LTB<sub>4</sub>, and LTD<sub>4</sub> were isocratically eluted in an HPLC system (Waters Associates) with a mixture of 72% methanol, 28% water (both HPLC grade), and 0.5 mmol/L ammonium acetate; acetic acid was added to obtain a pH of 5.88. Purified preparations of LTC<sub>4</sub>, LTD<sub>4</sub>, and LTB<sub>4</sub> (Merck Frost, Montreal, Canada) were used as standards and tested in parallel with the samples. The solvent for HPLC was pumped through an HPLC pump (501 HPLC, Waters), and the effluent was monitored at 280 nm with a spectrophotometer (Lambda MAX model 81 LC spectrophotometer, Waters). Data were processed with a Waters Baseline 810 chromatography computerized work station. The technician performing the leukotriene measurements was blinded to the type of treatment. The lower limit of detection of our assay was 0.5 ng/mL.

Our all-step recovery rates of standard leukotrienes include incubation with methanol/water, extraction with Sep-Pak cartridge, vacuum evaporation, redissolution, and passage through HPLC.

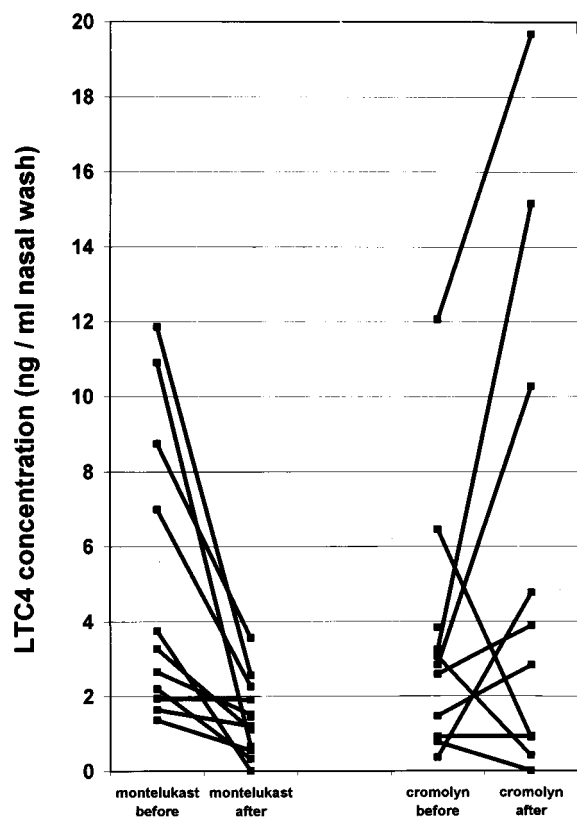


FIG 2. LTC<sub>4</sub> concentration (in nanograms per milliliter) in nasal washes of children before and after first month of treatment with montelukast or cromolyn.

### Measurement of ECP

ECP was measured by double-antibody RIA with the Pharmacy ECP RIA kit (Pharmacia and Upjohn Diagnostics, Uppsala, Sweden). The assay uses a specific antibody against human ECP raised in rabbit with iodine 125-labeled ECP as tracer. The assay was conducted in 50  $\mu$ L of nasal supernatant. The sensitivity of the assay was 2  $\mu$ L/L, and the within-assay variation was 11%.

### Statistical analysis

The statistical analysis was performed with use of BMDP statistical software.<sup>24</sup> Means and SEMs were computed for each of the continuous variables, and differences were compared with 1-way ANOVA and ANOVA with repeated measures. The significance for categorical variables was computed with use of Pearson's chi-square test as well as Fisher's exact test. A *P* value <.05 was considered significant.

### RESULTS

The results of 3 of the original 26 children who refused to continue with the collection of nasal washes were excluded from the analysis. Nasal samples were obtained before and after the first treatment period in the remaining 23 children. The cross-over analysis was performed only in the 17 children (9 given montelukast and 8 cromolyn) from whom nasal wash samples were obtained at all 4 stages of the study. Twelve children par-

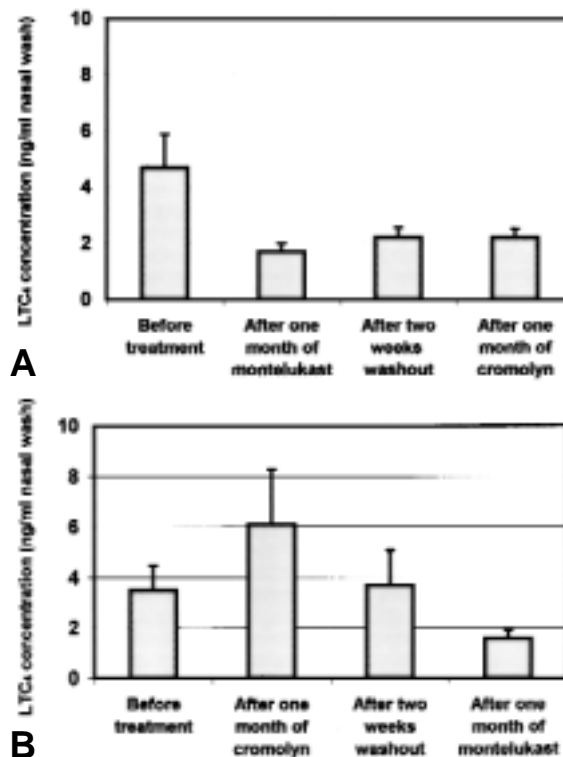


FIG 3. LTC<sub>4</sub> concentration in nasal washes of the children in all 4 periods of study. A, Children starting treatment with montelukast. B, Children starting treatment with cromolyn.

ticipated in the extension trial (6 were treated with montelukast and 6 with beclomethasone).

Twelve children started treatment with montelukast and 11 with cromolyn. The 2 groups were comparable for baseline data (Table I). Most of the children (74%) had been treated continuously with inhaled corticosteroids before the study. The mean prestudy peak expiratory flow rate of the whole cohort was 71% to 78% of predicted. Eighty-three percent of the children had a family history of asthma. During treatment with montelukast 75% of the children used fewer than 2 puffs of  $\beta_2$ -agonists (Ventolin) per week compared with 27% during treatment with cromolyn (*P* < .04).

During the entire follow-up period with montelukast, the children did not have an asthma exacerbation and they did not use any antiasthma drug except for  $\beta_2$  agonists and these only "as needed." However, 2 of the children treated with cromolyn withdrew from the study because of asthma exacerbation. When asked at the end of the study, 82% of the parents and 76% of the children preferred the treatment with montelukast, and only 2% of the parents and 27% of the children preferred cromolyn (the others had no preference). No adverse effects were noted in any of the children with either treatment.

All children treated first with montelukast showed a decrease in the LTC<sub>4</sub> concentration after 1 month of treatment, from a mean ( $\pm$  SE) of 5.03  $\pm$  1.17 ng/mL of nasal wash to 1.42  $\pm$  0.33 ng/mL (*P* < .005). By contrast,

the LTC<sub>4</sub> concentration in the children receiving cromolyn rose although not significantly, after the first month from 3.37 ± 1.11 ng/mL before treatment to 5.88 ± 2.17 ng/mL after (*P* = .17) (Fig 2).

In the children who started treatment with montelukast, the LTC<sub>4</sub> concentration did not return to the pretreatment level at the end of the washout period (Fig 3, A). In the children who started with cromolyn, the LTC<sub>4</sub> concentration returned to the pretreatment level at the end of the washout period and then further decreased when montelukast was administered (Fig 3, B). The LTC<sub>4</sub> concentration of all the children (from both arms of the cross-over study) was significantly lower during the month in which they were treated with montelukast than in the month they received cromolyn (1.58 ± 0.23 vs 4.09 ± 1.21 ng/mL, *P* = .02) (Fig 4).

The recovery rate of standard LTC<sub>4</sub> including all the separation steps was 40%, whereas the rates for LTD<sub>4</sub>, LTB<sub>4</sub>, and LTE<sub>4</sub> were 15%, 10%, and 0%, respectively.

LTD<sub>4</sub> was detected only in 9 of 23 of the children before the study, in 5 of 23 after 1 month of treatment with montelukast, and in 8 of 23 after 1 month of treatment with cromolyn. The small number of patients with detected LTD<sub>4</sub> did not allow for statistical evaluation.

During the extension trial 6 children received montelukast and 6 beclomethasone. The mean concentration of LTC<sub>4</sub> in the children treated with montelukast remained low and was lower than the concentration found in the children treated with beclomethasone after 3 months (0.8 ± 0.7 vs 1.5 ± 0.6 ng/mL, *P* = .12) and after 6 months (1.0 ± 0.3 vs 1.5 ± 1.0 ng/mL, *P* = .26). The difference was not statistically significant, possibly because of the small number of patients.

The concentration of ECP decreased after the first month of treatment in most nasal washes of the children given montelukast (9/12 children), from a mean (±SE) of 284 ± 15 ng/mL of nasal wash to 241.3 ± 23 ng/mL (*P* = .12). By contrast, a small change in the opposite direction was noted in the children receiving cromolyn: 246 ± 20 ng/mL before and 260 ± 28 ng/mL after treatment. There was a borderline significant interaction between treatment group and effect of treatment (*P* = .08).

## DISCUSSION

This study demonstrates that treatment with montelukast, a leukotriene receptor antagonist, is associated with a significant suppression of LTC<sub>4</sub> release in the respiratory tract of children and with concomitant clinical improvement in their asthma status. These pathophysiologic and clinical effects were not observed when the same children received cromolyn therapy.

The ideal model for the investigation of mediator metabolism in asthma is the respiratory mucosa of the lung by either bronchial biopsy specimen study or by segmental BAL.<sup>19</sup> However, because these procedures are invasive, several groups have used the nose to study the underlying mechanisms of allergic and nonallergic reactions in the respiratory tract.<sup>25,26</sup> The nose allows for

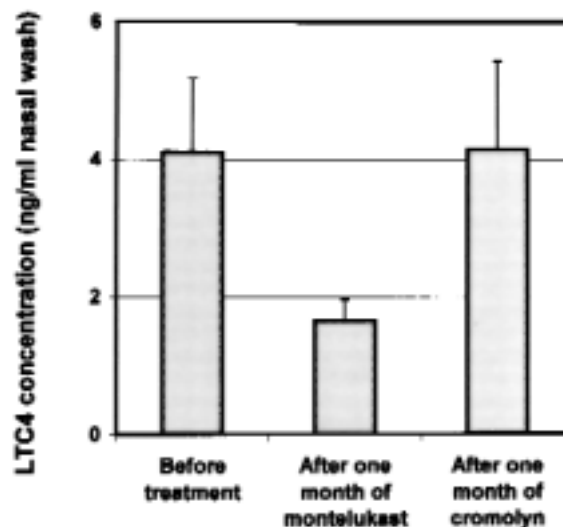


FIG 4. LTC<sub>4</sub> concentration in all the children (both arms of cross-over study) before and after treatment with montelukast or cromolyn.

the repetitive collection of nasal secretions after different interventions. In a trial with tracheotomized children,<sup>18</sup> we demonstrated a positive correlation between the concentration of leukotrienes in the bronchial tree (trachea) and in the nose.

LTC<sub>4</sub> was detected in all the samples at baseline and in most of the samples during the other treatment periods of the study, whereas LTD<sub>4</sub> and LTB<sub>4</sub> were detected in fewer samples. Studies have shown that LTC<sub>4</sub> is the principal leukotriene secreted by activated eosinophils.<sup>27</sup> Wenzel et al<sup>14</sup> reported that LTC<sub>4</sub> was the predominant sulfidopeptide leukotriene found in the BAL fluid of patients with atopic asthma undergoing allergen challenge, followed by LTD<sub>4</sub> and LTE<sub>4</sub>. In previous studies we demonstrated the presence of LTC<sub>4</sub> in the nasal washes of children with acute viral infection,<sup>15</sup> with bronchiolitis caused by respiratory syncytial virus infections,<sup>16</sup> and after exposure to ragweed antigen.<sup>17</sup> We have detected LTC<sub>4</sub> in tracheal secretions,<sup>18</sup> ear fluid,<sup>21</sup> and segmental BAL fluid.<sup>19</sup> The results of the current study indicate that LTC<sub>4</sub> is present in the respiratory mucosa of children with persistent asthma and that an active leukotriene receptor antagonist such as montelukast can suppress these elevated levels.

The exact mechanisms underlying the suppression of LTC<sub>4</sub> in the nasal washes of the children treated with montelukast are unclear. We offer 2 possible explanations. (1) Both LTC<sub>4</sub> and LTD<sub>4</sub> are known to share the same cysteinyl leukotriene T<sub>1</sub> receptor (CysLT<sub>1</sub>).<sup>28</sup> Cysteinyl leukotrienes promote eosinophil recruitment in the airways,<sup>29</sup> and eosinophils in turn, release leukotrienes.<sup>28</sup> Montelukast, which is a CysLT<sub>1</sub> receptor antagonist,<sup>28</sup> indirectly leads to a decrease in leukotriene concentration in the airways by reducing the influx of eosinophils. During the active asthma state (end of the run-in period), when the first baseline samples were obtained, the con-

centrations of leukotrienes in the respiratory mucosa were elevated. Immediately after treatment with montelukast, there may still have been a transient accumulation of leukotrienes, but they are unstable in the respiratory mucosa and undergo rapid metabolism and degradation.<sup>29</sup> Once they were absent from the airways, all the clinical effects of the leukotrienes,<sup>30</sup> including the attraction of eosinophils into the lung,<sup>31</sup> were prevented, ultimately resulting in a reduction in LTC<sub>4</sub> production. This theory is supported by the relative decrease in ECP (produced by eosinophils) observed in our study after treatment with montelukast. (2) The reduction in leukotrienes in the mucosa during treatment with montelukast may also be associated with another mechanism. LTD<sub>4</sub> increases nasal mucosal blood flow.<sup>32</sup> A montelukast-induced reduction in the concentration of leukotrienes may result in a reduction in blood flow into the respiratory mucosa, leading to a lower number of eosinophils arriving from the blood to the nose and thereby a reduced production of LTC<sub>4</sub>.

We used a cross-over design because the study was part of a "preference" international multicenter cross-over trial (not a clinical comparison of the efficacy and safety of the drugs). The main objective of the current study was to verify that montelukast, and not other drugs, induces a reduction in leukotriene concentration in the respiratory mucosa. The results for the group that started with montelukast and continued with cromolyn showed that the mean concentration of LTC<sub>4</sub> at the end of the washout period remained as low as that during active treatment with montelukast and did not reach pretreatment values. This suggests that montelukast had a cross-over effect during the 2-week washout period. We therefore performed both a cross-over analysis and a direct comparison of pretreatment and posttreatment values in the 2 parallel groups. The results proved to be comparable, demonstrating a significant reduction in LTC<sub>4</sub> concentration only during treatment with montelukast and not with cromolyn.

The scarcity of leukotrienes in the human body fluid, their rapid metabolism and degradation once they are formed in the respiratory mucosa,<sup>31</sup> and the cross-reactivity within the group of leukotrienes LTC<sub>4</sub>, LTD<sub>4</sub>, and LTE<sub>4</sub> in RIAs<sup>23</sup> necessitate the use of refined laboratory techniques, including separation and purification of the samples to detect and measure their concentration with HPLC.

The recovery rate of standard LTC<sub>4</sub> observed here with our separation technique was undesirably low. Measurement of the recovery rate of only 1 step of the procedure might have yielded a result closer to that in our previous study (76%),<sup>15</sup> with the same technique, which was similar to the 77% reported by others for the same separation step.<sup>8</sup> As such, there are 2 main steps in our procedure that might account for the relatively low recovery rate obtained in the current study: (1) Sep-Pak extraction, which precedes HPLC, is associated with a loss of leukotriene concentration and hence a low recovery rate<sup>34</sup> and (2) vacuum extraction and redissolution of the

leukotrienes from the dried tube always leaves some leukotrienes on the wall of the assay tube. As in our study, Schwartzberg et al<sup>33</sup> also reported a low recovery rate for LTD<sub>4</sub> that was less than half of that for LTC<sub>4</sub>. We did not detect LTE<sub>4</sub> mainly because our HPLC separation technique was optimally adjusted to a good separation of LTC<sub>4</sub> and LTD<sub>4</sub>.

The preliminary results of the extension part of the trial indicate that the concentration of leukotrienes in the respiratory mucosa remains low during 3 and 6 months of treatment with montelukast. The LTC<sub>4</sub> concentration was lower in the children treated with montelukast than in those treated with beclomethasone, indicating the specificity of the treatment with the antileukotriene agent.

A recent study had shown that cromolyn only attenuates, without statistical significance, the release of LTC<sub>4</sub> and LTD<sub>4</sub> in the nasal airway of allergic patients undergoing nasal allergen provocation.<sup>34</sup> In our study, treatment with cromolyn did not induce any significant change in the LTC<sub>4</sub> concentration, as opposed to treatment with montelukast. On the contrary, children treated with cromolyn showed an increase (although not statistically significant) in the concentration of leukotrienes in the respiratory mucosa. This may have been the result of deterioration of the clinical and pathophysiologic status of most of these children, who had been previously treated with inhaled corticosteroids.

Treatment with other antileukotriene drugs in humans, such as zileuton, a 5-lipoxygenase inhibitor,<sup>4,10</sup> or MK-0591, a 5-lipoxygenase activating protein inhibitor,<sup>5</sup> has been associated with the inhibition of urinary excretion of LTE<sub>4</sub>. Our study is the first to demonstrate a reduction in LTC<sub>4</sub> in the respiratory tract of children in response to a leukotriene receptor antagonist.

The current study demonstrates that montelukast causes a reduction in the concentration of leukotrienes in the respiratory mucosa of children with persistent asthma parallel to reduction in ECP and clinical improvement. This effect was not observed when the same children were treated with cromolyn.

We thank Mrs Gloria Ganzach for editorial assistance.

## REFERENCES

- Orning L, Kaijser L, Hammarstrom S. In vivo metabolism of leukotriene C<sub>4</sub> in man. *Biochem Biophys Res Commun* 1985;130:214-20.
- Drazen JM, Israel E. Asthma: a solution of half the puzzle? *Am Rev Resp Dis* 1991;144:743-4.
- Kikawa Y, Hosoi S, Inoue Y, Saito M, Nakai A, Shigematsu Y, et al. Exercise-induced urinary excretion of leukotriene E<sub>4</sub> in children with atopic asthma. *Pediatr Res* 1991;29:455-9.
- Hui KP, Taylor JK, Taylor GW, Rubin P, Kesterson J, Barnes NC, et al. Effect of a 5-hydroxygenase inhibitor on leukotriene generation and airway response after allergen challenge in asthmatic patients. *Thorax* 1991;46:184-9.
- Diamant Z, Timmers MC, ver der Veen H, Friedman BS, De-Smet M, Depre M, et al. The effect of MK-0591, a novel 5-hydroxygenase activating protein inhibitor, on leukotriene biosynthesis and allergen-induced airway responses in asthmatic subjects in vivo. *J Allergy Clin Immunol* 1995;95:42-51.
- Westcott JY, Smith HR, Wenzel SE, Larson GL, Thomas RB, Felsien D,

- et al. Urinary leukotriene E<sub>4</sub> in patients with asthma: effect of airways reactivity and sodium cromoglycate. *Am Rev Res Dis* 1991;143:1322-8.
7. Chavis C, van Vyve T, Chanez P, Farce M, Bousquet J, Michel FB, et al. Leukotriene E<sub>4</sub> plasma levels in adult asthmatic patients with variable disease severity. *Allergy* 1997;52:589-92.
  8. Creticos PS, Peters SP, Adkinson F Jr, Naclerio RM, Hayes ED, Norman PS, et al. Peptide leukotriene release after antigen challenge in patients sensitive to ragweed. *N Engl J Med* 1984;310:1626-30.
  9. Lam S, Chan H, LeRiche JC, Chan-Yeung M, Salari H. Release of leukotrienes in patients with bronchial asthma. *J Allergy Clin Immunol* 1988;81:711-7.
  10. Israel E, Rubin P, Kemp JP, Grossman J, Pierson W, Siegal SC, et al. The effect of inhibition of 5-hydroxygenase by zileuton in mild to moderate asthma. *Ann Intern Med* 1993;119:1059-66.
  11. Kikawa Y, Miyanomae T, Inoue Y, Saito M, Nakai A, Shigematsu Y, et al. Urinary leukotriene E<sub>4</sub> after exercise challenge in children with asthma. *J Allergy Clin Immunol* 1992;89:1111-9.
  12. Shindo K, Fukumura M, Miyakawa K. Plasma levels of leukotriene E<sub>4</sub> during clinical course of bronchial asthma and the effect of oral prednisolone. *Chest* 1994;105:1033-41.
  13. Wardlaw AJ, Hay H, Cromwell O, Collins JV, Kay AB. Leukotrienes, LTC<sub>4</sub> and LTB<sub>4</sub>, in bronchoalveolar lavage in bronchial asthma and other respiratory diseases. *J Allergy Clin Immunol* 1989;84:19-26.
  14. Wenzel SE, Larson GL, Johnston K, Voelkel NF, Westcott JY. Elevated levels of leukotriene C<sub>4</sub> in bronchoalveolar lavage fluid from atopic asthmatics after endobronchial allergen challenge. *Am Rev Respir Dis* 1990;142:112-9.
  15. Volovitz B, Faden H, Ogra PL. Release of leukotriene C<sub>4</sub> in the human respiratory tract during viral infection. *J Pediatr* 1988;112:218-22.
  16. Volovitz B, Welliver R, DeCastro G, Krystofik DA, Ogra PL. The release of leukotrienes in the respiratory tract during infection with respiratory syncytial virus: role in obstructive airway disease. *Pediatr Res* 1988;24:504-6.
  17. Volovitz B, Osur S, Bernstein JM, Ogra PL. Leukotriene C<sub>4</sub> (LTC<sub>4</sub>) release in respiratory mucosa during natural exposure to ragweed in ragweed-sensitive children. *J Allergy Clin Immunol* 1988;82:414-8.
  18. Volovitz B, Nathanson I, DeCastro G, Kikuawa T, Mukherjee A, Brodsky L, et al. Relationship between leukotriene C<sub>4</sub> and a uteroglobin-like protein in nasal and tracheobronchial mucosa in children: implication in acute respiratory illnesses. *Int Arch Allergy Appl Immunol* 1988;86:420-5.
  19. Sedgwick JB, Calhoun WJ, Gleich GJ, Kita H, Abrams JS, Schwartz LB, et al. Immediate and late airway response of allergic rhinitis patients to antigen challenge: characterization of eosinophil and mast cell mediators. *Am Rev Res Dis* 1991;144:1274-81.
  20. Iwasaki E. Leukotriene C<sub>4</sub> in children with atopic asthma. I: plasma levels in acute asthma. *Acta Paediatr Jpn* 1989;31:286-94.
  21. Brodsky I, Faden H, Bernstein J, Stanievich J, DeCastro G, Volovitz B, et al. Arachidonic acid metabolites in middle ear effusions of children. *Oto Rhinol Laryngol* 1991;100:589-92.
  22. Reiss TF, Chervinsky P, Dockhorn RJ, Shingo S, Scidenberg B, Edwards TB. Montelukast, a once-daily leukotriene receptor antagonist in the treatment of chronic asthma. *Arch Intern Med* 1998;158:1213-20.
  23. Knorr B, Matz J, Bernstein JA, Ngugen H, Seidenberg BC, Reiss TF, et al. Montelukast for chronic asthma in 6-to 14-year-old children: a randomized, double-blind trial. *JAMA* 1998;279:1181-6.
  24. BMDP statistical software. Dixon WJ, editor. Los Angeles: University of Californian Press; 1990.
  25. Picado C, Ramis I, Rosello J, Prat J, Bulbena O, Plaza V, et al. Release of peptide leukotriene into nasal secretions after local instillation of aspirin in aspirin-sensitive asthmatic patients. *Am Rev Respir Dis* 1992;145:65-9.
  26. Naclerio RM, Meier HL, Kagey-Sobotka, Adkinson NF, Meyers DA, Norman PS, et al. Mediator release after nasal airway challenge with allergen. *Am Rev Respir Dis* 1983;128:587-602.
  27. Weller PF, Lee CW, Foster DW, Curey EJ, Austen KF, Lewis RA, et al. Generation and metabolism of 5-lipoxygenase pathway leukotrienes by human eosinophils: predominant production of leukotriene C<sub>4</sub>. *Proc Natl Acad Sci U S A* 1983;80:7626-30.
  28. Kumlin M, Dahlen SE. Characteristics of formation and further metabolism of leukotrienes in the chopped human lung. *Biochem Biophys Acta* 1990;1044:201-10.
  29. Laitinen LA, Laitinen A, Haahtela T, Vikka V, Spur BV, Lee TH. Leukotriene E<sub>4</sub> and granulocytic infiltration into asthmatic airways. *Lancet* 1992;341:989-90.
  30. Smith LJ, Greenberger PA, Patterson R, Krall RD, Bernstein PR. The effect of inhaled leukotriene D<sub>4</sub> in humans. *Am Rev Respir Dis* 1985;131:368-72.
  31. Owen WF Jr, Soberman RJ, Yoshimoto T, Sheffer AL, Lewis RA, Austen KF. Synthesis and release of leukotriene C<sub>4</sub> by human eosinophils. *J Immunol* 1987;138:532-8.
  32. Bisgaard H, Olsson P, Bende M. Effect of leukotriene D<sub>4</sub> on nasal mucosal blood flow, nasal airway resistance and nasal secretion in humans. *Clin Allergy* 1986;16:289-97.
  33. Schwartzberg SB, Shelov SP, van Praag D. Blood leukotriene levels during the acute asthma attack in children. *Prostaglandins Leukotrienes Med* 1987;26:143-55.
  34. Ramis I, Catafau JR, Serra J, Bulbena O, Picado C, Gelpi E. In vivo release of 15-HETE and other arachidonic acid metabolites in nasal secretions during early allergic reactions. *Prostaglandins* 1991;42:411-20.